# Operation Manual Shodex GPC K-2000 series

(Please read this operation manual carefully to achieve the best and consistent column performance for a long time.)

### **Important Handling Instructions**

### Caution!

 Please consult the Safety Data Sheet (SDS) of reagents and solvents used with the column and understand their proper handling methods to prevent potential health hazards or death from occurring.

• Please wear appropriate personal protective equipment such as lab goggles and gloves when handling organic solvents and acid and alkaline reagents. Avoid any direct physical contact to prevent chemical injuries.

### **Before Using the Column**

- (1) Please visually inspect the column package and the column surface for any damage.
- (2) Please check the product name and serial number (Serial no. or S/N) written on the column package and adhesive label on the column body.
- (3) Please download the Certificate of Analysis (CoA) for the purchased product. The CoA can be downloaded from Shodex website (https://www.shodex.com/download/). You will be asked to enter the serial number.

#### 1. Introduction

Thank you for purchasing the Shodex product. Shodex GPC K-2000 series is size exclusion chromatography columns used with organic solvents. The column series is suitable for preparative isolation of organic compounds and oligomers. The columns with different exclusion limits are available. Please select columns with target molecular weight ranges (see below) that meet the molecular weight range of your samples.

### 2. Column Components

Please refer to the Shodex website: https://www.shodex.com/en/da/01/07.html

	Column Size (mm)					
Product Code	Product Name	Column Size (mm)		Particle Size	Plate Number	Exclusion Limit <sup>*1</sup>
		I.D.	Length	(µm)	(Per Column)	
F6102301	GPC K-2001	20.0	300	6	≥ 18,000	1,500
F6102312	GPC K-2002	20.0	300	6	≥ 18,000	5,000
F6102315	GPC K-2002.5	20.0	300	6	≥ 18,000	20,000
F6102303	GPC K-2003	20.0	300	6	≥ 18,000	70,000
F6102304	GPC K-2004	20.0	300	7	≥ 14,000	400,000
F6102305	GPC K-2005	20.0	300	10	≥ 10,000	4,000,000
F6102306	GPC K-2006	20.0	300	10	≥ 10,000	(20,000,000) <sup>*2</sup>
F6102309	GPC K-2006M	20.0	300	10	≥ 10,000	(20,000,000)*2
F6700407	GPC K-G 8B	8.0	50	15	(Guard Column)	-

### 3. Column Specifications

K-2006M is a mixed-gel column, provides wider molecular-weight analysis ranges.

<sup>\*1</sup> Reference value only / Measured with polystyrene <sup>\*2</sup> Estimated value

Base Material : Spherical porous particles of styrene divinylbenzene copolymer

Column Housing : SUS-316

Screw Type : Internally-threaded type No.10-32 UNF

Shipping Solvent : Chloroform

### 4. Usable Conditions

## 4.1 System Settings

Product Name	Flow Rate (	mL/min)	Maximum Pressure (MPa/column)	Temperature (°C)	
	Recommended	Maximum		Recommended	Maximum
GPC K-2001 to K-2006M	3.5	4.5	2.0	RT* - 40	60
GPC K-G 8B	-	-	-	KT - 40	00

\*Room temperature

## 4.2 List of Applicable Solvents

	Product Name					
Solvents	K-2001	K-2002, K-2002.5	K-2003	K-2004 to K-2006, K-2006M		
Tetrahydrofuran(THF)	✓	✓	✓	<ul> <li>✓</li> </ul>		
Benzene	✓	✓	✓	×		
Toluene	✓	✓	✓	✓		
Carbon tetrachloride	×	✓	✓	✓		
p-Xylene	×	✓	✓	×		
Dioxane	×	✓	✓	✓		
30 % m-Cresol/Chloroform	×	✓	✓	✓		
30 % o-Chlorophenol/Chloroform	×	✓	✓	✓		
30 % HFIP/Chloroform	×	✓	✓	✓		
o-Dichlorobenzene (ODCB)	×	×	✓	✓		
1,2,4-Trichlorobenzene (TCB)	×	×	✓	✓		
Diethyl ether	×	×	✓	✓		
Ethyl acetate	×	×	✓	✓		
Acetone	×	×	✓	✓		
Methyl ethyl ketone	×	×	✓	×		
N,N-Dimethylformamide (DMF)	×	×	✓	✓		
N,N-Dimethylacetamide (DMAc)	×	×	✓	✓		
m-Cresol	×	×	✓	✓		
o-Chlorophenol	×	×	✓	✓		
Quinoline	×	×	✓	<ul> <li>✓</li> </ul>		
N-Methylpyrrolidone (NMP)	×	×	✓	✓		
Hexafluoroisopropanol (HFIP)	×	×	×	✓		
Dimethyl sulfoxide (DMSO)	×	×	×	×		
Hexane	×	×	×	×		
Acetonitrile	×	×	×	×		
Methanol	×	×	×	×		
Water	×	×	×	×		

✓: Can be used X: Cannot be used

The standard solvent is chloroform.

### Caution!

 Solvents such as chloroform that contains chlorine are toxic. Ventilate the room well when using them.

### Attention!

- Use the column within above stated flow rate, pressure, and temperature ranges. Using the column outside the given range may damage the column and lower its performance.
  - Column pressure is influenced by eluent composition, flow rate, and column temperature. When changing the eluent compositions, adjust the flow rate and column temperature so that the column pressure remains below the usable maximum pressure.
  - Shear degradation occurs more likely in larger molecular weight compounds. The result of shear degradation may appear as lower molecular weight measurement than the actual value and/or low reproducibility. If shear degradation is suspected, use a lower flow rate.

### 5. Eluent Preparation

- (1) Degas the eluent fully to prevent the formation of air bubbles.
- (2) Presence of small debris or insoluble substances may result in deterioration of columns and/or they may appear as noise on chromatograms. Filter the eluent with a 0.45-µm disposable filter to prevent the problems from occurring.

# Attention!

- 1. Use of HPLC grade organic solvents of guaranteed quality, which can be used without problems in HPLC is recommended. If organic solvents with different grades are used together, make sure that their qualities are all suitable for the analysis prior to the use. Solvents left in opened bottles for a long time should not be used. The content may have been changed, absorbed moisture, or has been contaminated.
  - · Always use freshly prepared solvents. Solvents stored for a long time may have changed their compositions and may influence elution patterns and/or damage the column.



· Use of an on-line degasser is recommended.

### 6. Sample Preparation

- (1) If possible, use the eluent for analysis to dissolve or dilute samples. If this is difficult, use a solvent which has a composition that is as close as possible to the eluent composition and which fully dissolves or dilutes the sample.
- (2) Filter diluted sample solutions using disposable 0.45-µm filters to prevent the column from clogging or deteriorating.
- (3) To prepare samples with molecular weights larger than 1,000,000, first allow the sample to stand in the eluent of analysis for 1 day until it becomes fully swollen. Next, slowly agitate the sample solution to completely dissolve the sample. Be careful as aggressive agitation can cut the polymer chains of the analyte.
- (4) Suggested injection volume is 250 to 1,000  $\mu$ L.
- (5) Viscosity of high molecular weight compound is largely influenced by its molecular weight and concentration. Samples with high viscosity cause peak broadening and elution delay, and this makes it difficult to obtain their accurate molecular weight distributions. In general, the larger the molecular weight of the compound, the higher its viscosity becomes. To suppress the influence from high viscosity, it is recommended to lower the sample concentration. Please use the below table as a reference when preparing samples for molecular weight distribution analyses.

Molecular Weight Range	Optimal Concentration (w/v)	
≤ 5,000	≤ <b>1.0</b> %	
5,000 - 25,000	$\leq$ 0.5 %	
25,000 - 200,000	≤ 0.25 %	
200,000 - 2,000,000	≤ 0.1 %	
≥ 2,000,000	≤ 0.05 %	

(6) General organic substances can be injected at a higher concentration than polymer. Suggested sample concentration is less than 10 % (w/v). Overload does not damage the column, but if the peak shape is distorted, lower the sample concentration.

(7) When sample contains water, methanol, or hexane, remove them prior to the injection.

- Attention! When a sample is dissolved in a solvent other than the eluent and if the sample matrix contains components which do not dissolve in the eluent fully, precipitates may form and clog the column.
  - Note

 $\cdot$  Use of a guard column is recommended to protect the analytical column.

### 7. Column Usage Procedure

#### 7.1 HPLC System Preparation

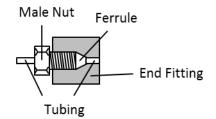
Wash the entire HPLC system prior to column installation, including all flow-lines and sample loop by switching the valves, and then replace the washing solution with the eluent to be used. If desired new eluent has low miscibility/solubility to the eluent of previous analysis, first use the eluent that is miscible/soluble to both eluents, and then replace it with the desired eluent.

# Attention!

- · If the eluent left in the HPLC system is not compatible with the column to be used, it may damage the column.
- $\cdot$  A drastic change in the eluent compositions may remove substances adsorbed on the HPLC system and they may enter and deteriorate the column.

### 7.2 Column Installation

- (1) Connect the column to HPLC system by following the "flow direction arrow" (→) indicated on the column adhesive label. If a guard column is used, position the guard column in front (before the inlet) of the analytical column.
- (2) Make sure to insert the tubing all the way to the end fitting and secure it with the male nut. It is important that there is no extra space between the tubing and the column side of the end fitting. Presence of an extra space will let the sample to spread out and may result in wide peaks.



- (3) Set the initial flow rate at less than 1.0 mL/min and start the system. If the column is to be heated during the analysis, keep the low flow rate until the column temperature reaches to the set temperature, and then gradually increase the flow rate to the desired temperature.
- (4) Multiple number of columns can be connected in series. When connecting multiple columns with different exclusion limits, set the column with higher exclusion limit at the upper stream position. When using K-2006M, use multiple number of K-2006M columns in series. This is because mixed-gel columns are prepared by using several different gels with different exclusion limits, designed to provide linear calibration curves over the set molecular weight ranges. If different columns were used together, this changes the mixing ratio of the gels and consequently it may deform the linear calibration curve.

# Caution!

· Verify that there is no solvent leak. The solvent leak may cause electronic leakage, rust, and/or chemical injury.

Attention!

- Make sure not to let air bubbles enter the column while installing the column. The air bubbles may damage the column.
  - When restarting the system after column installation or after holding the eluent flow, start the system at less than 1.0 mL/min. A rapid increase in pressure can damage the column.

If the column was heated during the analysis, lower the flow rate to less than 1.0 mL/min at the end of analysis. Then, turn off the column oven to let the column temperature returns to room temperature before stopping the pump. This is to prevent creating an empty space in the column, which deteriorates the column. Since if the pump was stopped while the eluent inside the column is still hot, the eluent volume decreases and creates an empty space when the eluent temperature decreases.

Note

· It is recommended to set the pump limiter to avoid exceeding the maximum pressure.

#### 7.3 Solvent Exchange

To replace the solvent, set the column temperature at 40 to 60 °C and set the flow rate at less than 1.5 mL/min. Recommended solvent volume to introduce at each step is 3 to 5 times of the column volume. If replacing the solvent at room temperature, set the flow rate at less than 1.0 mL/min.

- (1) Check miscibility/solubility of the desired new solvent and the solvent currently filled in the column.
- (2) When replacing the current solvent with a miscible solvent, first introduce 1:1 mixture of the current solvent and the new solvent, and then replace it with 100 % new solvent.
  - e.g. When replacing from chloroform to DMF, first run 1:1 mixture of THF and DMF and then introduce 100 % DMF.
- (3) When replacing the current solvent with a solvent with low miscibility/solubility to the current solvent, first use a solvent that is miscible/soluble to both solvents, and then replace it with the new solvent.
  - e.g. When replacing from chloroform to "DMF with LiBr", first follow the example in procedure (2), and then replace 100 % DMF with "DMF with LiBr".

# Attention!

· Frequent solvent replacement deteriorates the column, and thus not recommended.

### 7.4 Column Cleaning

Problems in peak shapes and elution time changes or elevated column pressure are often caused by the deposition of insoluble or adsorbing components from the sample/flow-line inside the column. These problems may be resolved by cleaning the column.

If a guard column is used with an analytical column(s), first remove the guard column and check the performance of the analytical column alone. If the problem is solved, most likely the cause was from the guard column. In this case, clean the guard column.

If the problem remains even after removing the guard column, clean both guard and analytical columns. Make sure to clean the guard and the analytical columns separately. In case multiple number of analytical columns are used together, wash them separately. During the column cleaning, disconnect the detector and collect the washing solution directly from the column outlet into a waste container (i.e., do not let the solution go through the detector).

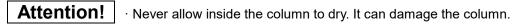
If the column performance does not improve (recover) after performing the column cleaning, please replace the column with a new one.

#### <Cleaning method>

Insoluble components that block the column inlet may be removed by reversing the flow direction, i.e., introducing the eluent from the column outlet, with flow rate at less than half of the recommended flow rate.

#### 8. Column Storage

Remove the column from HPLC system keeping the solvent used for the analysis. Securely tighten the end caps and store the column at a location with stable temperature (a cool and dark space is recommended). If the column is used with a solvent that may solidify during the storage (e.g., trichlorobenzene which has high melting point), replace the in-column solvent with the shipping solvent. Refer to section 7.3 Solvent Exchange for how to replace the eluent.



#### 9. Column Inspection

Please refer to the inspection method described in the CoA. At Shodex, "half width method" is adopted for the calculation of plate count and "asymmetry factor (Fas)" is adopted for the calculation of peak symmetry. Please refer to the Shodex website for the detail: https://www.shodex.com/en/da/07.html

Attention! • Plate count and Fas values change significantly depend on samples and/or analysis conditions being used. To check the initial column condition, please make sure to use the same sample and the analysis condition mentioned in the CoA.

#### 10. Additional Warnings

(1) Do not remove end fittings.

(2) Do not make a strong impact on the column. Do not drop or hit the column on a hard surface.

(3) Please follow a proper waste disposal method specified by your local regulations.

Please refer to the Shodex website (https://www.shodex.com/) for product details and their applications. For additional assistance, contact the distributor from whom you purchased the column or contact your regional Shodex support office (https://www.shodex.com/en/support\_office/list).